Exploring Unconventional Growing Methods to Promote Healthy Growth in the Common Household Plants *Tagetes patula* L. and *Lepidium sativum*

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#### **Abstract**

#### Chemical fertilizers have been used in increasing quantities for household gardening and commercial agriculture worldwide since their advent during the Green Revolution. Such fertilizers have detrimental impacts on the environment, contributing to nutrient runoff and aquatic dead zones. At the same time, water consumption for agricultural needs has skyrocketed. Alternative growing methods are urgently needed to reduce the impacts of plant cultivation. This study focused on finding more sustainable growing methods that reduce chemical fertilizer or water usage and can be used at the household level for garden plants. We hypothesized that the alternative growing methods would better encourage healthy plant growth as compared to a control. Several marigold (Tagetes patula L.) and garden cress (Lepidium sativum) plants were observed over a 13-week period. Metrics for healthy plant growth were height at first bloom, measured for each plant as the distance from the base to the tallest point; growing time, counted from day of sprout until the end of the experiment; and survival rate, calculated by dividing the number of surviving plants that bloomed by the number of plants that sprouted. The results indicated that the treatments did not have a statistically significant effect on marigold and garden cress growth times in addition to marigold heights. However, the deep water culture (DWC) treatment for garden cress plants significantly increased the height at first bloom compared to the control group. For survival rate, the treatments had little effect on the garden cress, but the eggshell grounds, wick system, and DWC system groups outperformed the control group for marigolds.

#### **Introduction**

Approximately 12,000 years ago, humanity developed agriculture, ending their formerly nomadic and hunter-gatherer lifestyle. Multiple factors led to this. The end of the last Ice Age set up favorable conditions for crops to grow. As a result of this, humanity began to harvest these plants, slowly transitioning to increasingly depend on and expand their fields. Permanent settlements developed around these agricultural sites, and civilizations prospered (1).

As agriculture has evolved, more complex techniques have been developed. For instance, slash-and-burn agriculture, the practice of burning areas of fertile land and allowing the ashes to resupply nutrients to the ground, has sustained crop yields for millennia (2). Later, improved techniques and tools developed, boosting productivity. For instance, crop rotation was used to improve the longevity and nutrient content of fields, and the seed drill reduced the amount of waste caused by avian predation on exposed seeds (3).

Most recently, the Green Revolution introduced modern technology into commercial and domestic agriculture, featuring chemical fertilizers and pesticides. Such chemical products have repercussions for the environment, namely increased phosphorous and nitrate concentrations from runoff (4). Excess nutrients enter bodies of water to create large algal blooms that can clog waterways and create “dead zones,” where aquatic species cannot thrive, due to the oxygen-consuming decomposition of dead algae (5).

In addition to affecting animal ecosystems, fertilizers also have the potential to directly harm humans. Furthermore, chemical runoff in waterways makes its way into fish, poisoning them and the humans that consume them (5). Nitrates from fertilizers may also easily enter the groundwater, poisoning surrounding communities and causing life-threatening diseases and conditions in humans due to the toxicity of elevated nitrate levels in the body (6). With approximately one-third of the world not having access to clean drinking water, the continued heavy application of chemical fertilizers presents an issue to human society (7). At the same time, global water usage for agricultural purposes such as crop irrigation has skyrocketed (8). To reduce the environmental impacts of plant cultivation, alternative growing methods that reduce chemical fertilizer use and water consumption are desperately required. One alternative growing method is the use of common household waste as sources of nutrients. Another alternative growing method involves substrate-based growing methods such as hydroponics. A study conducted by Blok, Jackson, et al. demonstrated higher growth yields from substrate-based growing methods, such as hydroponics, compared to soil-based growing methods due to superior transport rates of water, nutrients, and oxygen allowed by the former (8). Hydroponics involves growing plants in a non-soil substrate with a nutrient reservoir. Water from the nutrient solution is recirculated amongst plants and reused, thereby reducing its consumption.

We hypothesized that the application of alternative growing methods on the common garden plants dwarf French marigolds (*Tagetes patula* L.) and garden cress (*Lepidium sativum*) would better improve plant health, as determined by plant height at first bloom, plant survival rate, and growth time, in comparison to controls. We used common household items as sources of nutrients required by plants, which includes nitrogen, phosphorus, potassium, calcium, and magnesium (9). Human urine was used as an organic fertilizer for its high nitrogen content from its chief component, urea, and accessibility (10). Eggshells were another organic fertilizer, used due to their calcium content via calcium carbonate. Wood ash, produced from slash-and-burn agriculture and containing partially water-soluble calcium, potassium, and magnesium, was another selected fertilizer (2). We selected two common hydroponic systems for this project: wick and deep water culture (DWC). Wick systems use a wick to deliver the nutrient solution to the substrate while DWC systems partially submerge plant roots directly into the nutrient solution.

This experiment demonstrated how certain alternative growing methods can have beneficial effects on plant growth in terms of increasing plant height at first bloom, increasing plant survival rates, and decreasing plant growth time.

#### **Results**

The marigolds and garden cress were tested in the same manner. Germinated seeds were planted in soil or substrate, depending on the treatment. Group 1 (Control) seeds were grown conventionally in plain soil, Group 2 (Urea Solution) seeds were watered with a urine-based urea solution weekly, Group 3 (Burnt Foliage) seeds were given wood ashes weekly, Group 4 (Eggshell Grounds) were given finely-ground shards of eggshells weekly, Group 5 (Wick System) seeds were grown in a wick-style hydroponic system, and Group 6 (DWC System) seeds were grown in a deep water culture hydroponic system. Plant growth was monitored over a 13-week period.

First, the average plant height at first bloom was examined. For the marigolds, there was variation in average plant height, but there was no statistical significance between the six treatments (p = 0.058) (**Figures 1 & 2**). For the garden cress, there was variation in average plant height at first bloom as well. Plant height was significantly higher in Group 6 while lowest in Group 2 (p-value < 0.05, one-way ANOVA) (**Figures 1 & 2**). A post hoc Tukey-Kramer test with treated garden cress compared to the control revealed statistical significance only for the DWC treatment.

The effect of each treatment on the survival of each plant was also assessed. At the end of the 13-week experiment, the plant survival rate was calculated by dividing the number of plants continuing to grow after the first bloom by the number of plants that sprouted. For marigolds, the control group had the lowest rate plant survival, 0.4 (2 plants), while the eggshell grounds, wick system, and DWC System groups had the highest rate of plant survival, 1.00 (5 or 6 plants) (**Figures 1 & 3**). For garden cress, every treatment group had roughly equal rates of plant survival. The Urea Solution, Burnt Foliage, and Wick System groups had a survival rate of 0.83 (5 plants) while the control, eggshell grounds, and DWC System groups had a rate of 1.00 (6 plants) (**Figures 1 & 3**).

The average growth time, defined as the number of days the average plant took to bloom from the time of sprouting, was also assessed for each treatment. For the marigolds, there were no statistically significant differences between the average growth times for all six treatments (p = 0.077) (**Figures 1 & 4**). For the garden cress, there was a statistically significant difference in average growth time between treatments (p-value < 0.05, one-way ANOVA) (**Figures 1 & 4**). However, follow-up post hoc Tukey-Kramer tests between the control and other treatments revealed no statistical significance.

We also evaluated both plant varieties qualitatively. Starting in the twelfth week of experimentation, marigolds in Groups 2, 3, and 4 developed a purple color around the edges and tips of their lower leaves (**Figure 5**). There were no significant qualitative changes in any treatment groups of the garden cress plants.

#### **Discussion**

Select growing methods from this study encouraged healthy plant growth, as was shown by them yielding taller plants at first bloom, increasing survival rates, and decreasing growth times. The initial hypothesis was correct in some respects. This study demonstrated that simple, more environmentally-conscious growing methods can be applied to household gardens using common objects often overlooked in the house to reduce chemical fertilizer usage, as well as water-efficient hydroponic systems to reduce water consumption.

This study revealed that growing garden cress with a DWC hydroponics system increased garden cress height at first bloom at a statically significant level compared to the control treatment. However, the selected growing methods applied to marigolds did not result in significantly different plant heights at first bloom. As a result, it is not conclusive whether the selected methods have a significant effect on plant height at first bloom for marigolds. More data with other plant varieties is necessary to determine the efficacy of these methods or if these methods merely have a neutral effect on certain species. On the other hand, for the garden cress, the DWC treatment had the highest average height at first bloom, indicating its success in encouraging healthy plant growth. Taller plants, as compared to short plants, indicate the presence of critical nutrients and minerals required for healthy plant growth.

Although the eggshell grounds group for marigolds generated the highest average plant height at first bloom, there was no significant difference in height compared to the control group. This implies a possible lack in statistical power. Had there been more marigolds grown and observed, the results for plant height at first bloom might have been significant as power can be increased with a greater sample size.

The experiment also indicated that plant growth time was affected for only garden cress, though there was no significant difference between the control and other treatments. Longer growth times would indicate slower growth from a lack of nutrients, which is consistent with the findings. The lowest average growth time for garden cress came from the DWC treatment while the higher average growth times came from the in-ground treatments and control, which had fewer supplied nutrients. Shorter growth times would indicate heathy plant growth due to the presence of necessary nutrients. For marigolds, there was no meaningful difference between average growth times for all the treatments, demonstrating a difference in treatment efficacy based on plant types.

The discrepancy between the ANOVA results, which found statistical significance between the garden cress groups for growth time, and the post hoc Tukey-Kramer test, which found no statistical significance between the control and other treatments, is likely due to a lack of statistical power. Due to the small sample sizes, statistical power was low, thus reducing the chance a post hoc test would be able to detect a difference between treatments.

The study additionally demonstrates the impact each treatment has on the survival rate of both types of plants. While the survival rate for garden cress plants were roughly similar, the rates for marigolds varied. Group 1 marigolds (the Control group) had the lowest survival rate while Groups 4, 5, and 6 (the Eggshell Grounds, Wick System, and DWC System groups) had the highest survival rates. A higher survival rate indicates healthy plant growth as the soil contains the necessary nutrients. For the marigolds, the reason for a lower survival rate for the control group is obvious, as no nutrient-bearing treatment was applied. However, the higher survival rates for the Eggshell Grounds, Wick System, and DWC System treatments demonstrate their contribution to healthy plant growth since all were successful in sustaining every marigold that sprouted. For the cress, similar survival rates, where up to one plant died for each treatment, reveal the limited impact each treatment had in sustaining the garden cress plants.

Notably, across all the plant health metrics (height, survival rate, and time to bloom), DWC System groups consistently outperformed or were equal to other growing methods. This may point to the DWC System’s overall high capability of providing nutrients to plants. A similar case may be made for the Eggshell Grounds and Wick System groups for marigolds. However, in the case of the Burnt Foliage groups, this does not apply. In both marigolds and the garden cress, the plant survival rate remained the same, at a rate of approximately 0.8 with a similar rate of decline (dying off around the 40-day mark). This suggests that the burnt foliage may not be a very effective fertilizer. Similarly, Urea Solution groups across both plants had a similar survival rate; however, there was a greater disparity between when plant sprouts started to die off.

The development of a purple color in the leaves of marigolds in Groups 2, 3, and 4 indicate a phosphorus deficiency in the soil (11). As none of the treatments applied to the in-ground plants supplied phosphorus, this is not surprising. Further research may consider other unconventional growing methods to supplement the lack of phosphorus.

There are multiple limitations within this study. Only a maximum of 6 plants for each treatment were tracked over the 13-week period. Additionally, only two types of plants were taken into consideration for this experiment. Future studies should include a greater number of individual plants for each treatment. Though marigolds and garden cress, a flower and an herb, were selected to represent an average garden, a greater variety of garden plants such as fruits, vegetables, and legumes, which have different growing requirements, could be experimented on in further research. The narrow range of plant types also raises the question of whether these treatments would be practical on the wide-scale commercial agricultural level.

Other limitations of this study involve the manner in which it was conducted and the way the data was collected. This study was conducted for only 13 weeks. Further research may track these plants over months or years across different annual seasons and multiple growing seasons to evaluate the long-term effects of each treatment on the plants. Furthermore, this experiment only considered height at first bloom, survival rate, and growing time as metrics for plant health. Additional studies may consider more indicators of plant health such as weight and root condition. Moreover, plant height may be a slightly biased indicator due to natural variations in height that occur due to changing leaf positions at the time of daily data collection. Likewise, we only tracked height until the plant first bloomed. Changes in overall plant may have occurred after data collection stopped and may have therefore been missed.

#### **Methods**

# *Germination and planting*

To germinate the marigold and garden cress seeds, seeds were placed between moistened paper towels in an airtight plastic bag for 1–2 days at room temperature (70°F) until the seeds were hydrated. Once roots or sprouts began emerging, the seeds were transferred into the soil or growing medium at a depth of 0.25 inches (6 mm). Two marigold seeds and six garden cress seeds were planted in separate dirt pots. The soil used to plant marigold and garden cress seeds was taken from the same location. Two marigold seeds were planted in a 1:1 ratio of vermiculite and perlite for the Wick System treatment while only one seed was planted in perlite alone for the DWC treatment. Six garden cress seeds were planted in a 1:1 ratio of vermiculite and perlite for the Wick System treatment while only one seed was planted in perlite for the DWC treatment.

# *Plant maintenance*

Plants grown in dirt were watered daily. Each watering session thoroughly moistened the top of the soil. The plants were watered between two and six times daily, depending on the time of year and weather conditions. Watering was sufficient to keep plants from wilting. The marigold and garden cress pots and hydroponic systems were placed in an area with full sun. Any plants that died were removed and all weeds were removed upon detection.

# *In-ground treatment preparation and application*

To create the urea solution, samples of urine and tap water were combined into a 1:1 ratio. The solution was applied directly to the soil, enough to saturate the surface. To create the burnt foliage, dried lawn clippings and woodchips were burnt. About one teaspoon of ashes were spread on the surface of the soil. To create the eggshell grounds, several eggshells were washed and then ground into fine shards using a mortar and pestle. About 1 teaspoon of grounds was spread on the surface of the soil.

# *Hydroponics systems construction*

To create the Wick System hydroponics, a plastic juice bottle was rinsed and cleaned before being cut along its neck. The top of the bottle was inverted to serve as a pot. The bottom of the bottle was filled with the nutrient solution (detailed below). The wicks were created from recycled socks cut and plugged tightly into the bottle opening. Each wick clearly extended into the nutrient solution. The growing medium was a 1:1 mixture of perlite and vermiculite filling the pot to approximately 80% capacity (**Figure 6**).

To create the DWC System hydroponics, 6 oz. yogurt containers were recycled into a nest pots. Four slits longitudinally were cut in each container and four openings were made at the bottom corners (**Figure 7**). Twelve holes the size of a nest pot were cut into a block of Styrofoam large enough to fit into the top of the container. Each fashioned nest pot was filled with perlite to about 75% capacity. The block was made to float on top of the nutrient solution. At the bottom of the container, two air stones were attached on each side. Both air stones were attached to an air pump by plastic tubing and connectors (**Figure 8**).

# *Hydroponics maintenance*

All hydroponically grown plants were given a nutrient solution composed of water and plant food. The wick system plants were grown in a solution composed of 0.25 cup of water and 0.5 teaspoons of plant food. The DWC system plants were grown in a solution composed of 5 gallons of water and 7 teaspoons of plant food. Each week, the nutrient solutions were drained and replaced. Any occurrences of algae were removed with a 3% hydrogen peroxide solution and thoroughly rinsed with water.

# *Data collection*

Each day, the height of every plant was measured, from base of stem to the highest point of the plant (bud, flower, or leaf) using a length of string and ruler. The height at first bloom (when a bud opened into a flower) was used for statistical analysis. The plants were allowed to grow in a Southern California climate during the months of May to August for up to 13 weeks. Once a plant bloomed, researchers stopped tracking its plant height and observed only qualitative changes.

#### **References**

1. History.com Editors. “Neolithic Revolution.” *History.com*, A&E Television Networks, 12 Jan. 2018, www.history.com/topics/pre-history/neolithic-revolution.
2. Hauser, Stefan, and Lindsey Norgrove. “Slash-and-Burn Agriculture, Effects Of.” *Encyclopedia of Biodiversity*, 2013, vol. 2, pp. 551–562., doi:10.1016/b978-0-12-384719-5.00125-8.
3. The Editors of Encyclopaedia Britannica. “Agricultural Revolution.” *Encyclopædia Britannica*, Encyclopædia Britannica, Inc., 4 Dec. 2015, www.britannica.com/topic/agricultural-revolution.
4. Eghball, B., et al. “Long-Term Manure and Fertilizer Application Effects on Phosphorus and Nitrogen in Runoff.” *Transactions of the ASAE*, vol. 45, no. 3, 2002, doi:10.13031/2013.8850.
5. “How Fertilizers Harm Earth More Than Help Your Lawn.” *Scientific American*, Scientific American, 20 July 2009, www.scientificamerican.com/article/how-fertilizers-harm-earth/.
6. Ward, M., Jones, et al. “Drinking Water Nitrate and Human Health: An Updated Review.” *International Journal of Environmental Research and Public Health,* vol. 15, no. 7, 2018, doi:10.3390/ijerph15071557.
7. Boano, Fulvio, et al. “A Review of Nature-Based Solutions for Greywater Treatment: Applications, Hydraulic Design, and Environmental Benefits.” *Science of The Total Environment*, vol. 711, 2019, doi:10.1016/j.scitotenv.2019.134731.
8. Blok, Chris, et al. “Maximum Plant Uptakes for Water, Nutrients, and Oxygen Are Not Always Met by Irrigation Rate and Distribution in Water-Based Cultivation Systems.” *Frontiers in Plant Science*, vol. 8, 2017, doi:10.3389/fpls.2017.00562.
9. White, P. J., and P. H. Brown. “Plant Nutrition for Sustainable Development and Global Health.” *Annals of Botany*, vol. 105, no. 7, 2010, pp. 1073–1080. doi:10.1093/aob/mcq085.
10. Marieb, Elaine, and Katja Hoehn. *Human Anatomy & Physiology*. 8th ed., Benjamin Cummings, 2010.
11. Raese, J. Thomas. “Phosphorus Deficiency Symptoms in Leaves of Apple and Pear Trees as Influenced by Available Soil Phosphorus.” *Communications in Soil Science and Plant Analysis*, vol. 33, no. 3-4, 2002, pp. 461–477., doi:10.1081/css-120002757.

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#### **Data Figures**

1. **Descriptive statistics for marigolds and garden cress for three plant health metrics.** For both plants, the mean, standard deviation, and standard error for height at first bloom and growth time were calculated. The survival rate for each plant was also calculated.
2. **Figure 2: Average height at first bloom for marigolds** (left) **and garden cress** (right). Height was recorded when the plant first bloomed. There was no statistically significant difference between average heights at first bloom for all 6 marigold groups. A one-way ANOVA test was used to determine statistical significance, finding p > 0.05. There was a statistically significant difference between average heights at first bloom between the six garden cress groups. A one-way ANOVA test was used to determine statistical significance, finding p < 0.05. A post hoc Tukey-Kramer test revealed a statically significant difference between the control and DWC groups. An asterisk indicates the treatment with statistical significance compared to the control group. Error bars represent one standard error above and below the mean.
3. **Survival rate for marigolds** (top) **and garden cress** (bottom). The proportion of surviving plants was calculated by dividing the amount of plants that bloomed remaining at the end of the experiment over the amount of plants that sprouted. Plants that died during the experiment or never bloomed were not included. The survival rates from the marigolds is highest for the Eggshell Grounds, Wick System, and DWC System treatments (1) and lowest for the Control group (0.4). The survival rates for the garden cress are highest for the control group and Eggshell Grounds and DWC system treatments (all plants surviving) while the remaining treatments had one plant die.
4. **Average growth time for marigolds** (left) **and garden cress** (right). Growth time was recorded when the plant first sprouted until it bloomed. Plants that died during the experiment or never bloomed were not included. There was no statistically significant difference between average growth times for all six marigold groups. A one-way ANOVA test was used to determine statistical significance, finding p > 0.05. There was a statistically significant difference between average growth times between the six garden cress groups. A one-way ANOVA test was used to determine statistical significance between groups, finding p < 0.05. A post hoc Tukey-Kramer test, however, revealed no statically significant difference between the control and other treatment groups. Error bars represent one standard error above or below the mean.
5. **Representative photographs of the marigold plants with lower leaves that acquired a purple color**. Certain marigold plants in the Urea Solution, Burnt Foliage, and Eggshell Grounds groups developed a purple hue around the edges and tips of leaves lowest on their stems.
6. **Wick system set-up**. No plants are in the system for photographic purposes. A cleaned juice bottle was cut with the top inverted to use as a pot and the bottom as the nutrient reservoir. The growing medium is composed of a 1:1 ratio of perlite and vermiculite, which filled each pot to about 80% capacity.
7. **Nest pot construction and DWC system set-up**. No plants are in the nest pot for photographic purposes. A 6 oz. yogurt cup was recycled into a net pot by cutting slits into the sides. Perlite filled each nest pot to about 75% capacity.
8. **DWC system set-up**. No plants are in the system for photographic purposes. The DWC treatments for both plants were housed in a 7-gallon container. Twelve nest pots were placed into a block of Styrofoam and floated atop the nutrient solution.